

Overcoming Poor Germination in Australian Daisies

Kerry V. Bunker

Redlands Greenhouses Holdings, 191 Gordon Road, Redland Bay, Qld, 4165

INTRODUCTION

The evaluation and commercialisation of many Australian daisies has been limited by poor germination (Schaumann et al., 1987). Achenes of the Asteraceae consist of an embryo encased in a membranous coat (testa) which is surrounded by a fibrous outer coat (pericarp) which often has pappus hairs to aid dispersal. Both the testa and the pericarp have been identified as barriers to germination with exotic members of the Asteraceae. Removal of these layers, puncturing them, or soaking seeds in various solutions such as gibberellic acid are all reported to improve germination although the result is generally species specific (Atwater, 1980; Taylorson and Hendricks, 1977). Investigations were conducted to test the effects of scarification of the testa and pericarp, gibberellic acid, and light, on germination and dormancy of selected Australian daisies.

MATERIALS AND METHODS

Achenes from 27 species of Australian daisies were collected from wild populations throughout Western Australia and Queensland during the Summer of 1990 and 1991 (Table 1).

Achenes were stored at room temperature and germination trials conducted in petri dishes, within 36 weeks of collection. Germination trials were completely randomised designs of three germination treatments—intact achenes moistened with water (control), scarified achenes moistened with water, and intact achenes moistened with 500 mg liter⁻¹ GA₃—with two light levels (light and dark). There were five replicate petri dishes of 15 achenes for each species evaluated. Moistening solutions also contained 0.2% Thiram[®] fungicide (Rhone-Poulenc) and were applied at 5 ml per dish. Achenes were scarified by exposing a portion of the embryo with a dissecting needle. Petri dishes were placed on laboratory benches in ambient conditions. Mean daily minimum and maximum temperatures were 13±2C and 24±1C, respectively. Dark treatments were covered with aluminium foil while light treatments were exposed to 45±6 μM m⁻² s⁻¹ from a combination of fluorescent and natural light, for 10 to 12 h daily.

Germinated achenes were counted at Day 15 in dark treatments and every 3 days for 30 days in light treatments. At the end of the assessment period, achenes were dissected and those which contained undeveloped or no embryos were scored as nonviable. Germination was then recorded as percent viable achenes. Time (days) to 50% maximum germination (T₅₀) of intact achenes, following imbibition with water and exposure to light, was derived from plotted curves of mean percent germination against time for each species.

RESULTS AND DISCUSSION

Species responded differently to the treatments applied and germination following one or more treatments occurred in 17 of the 27 species evaluated (Table 1). The

testa and pericarp were influential in suppressing germination of *Leucochrysum stipitatum*, *Rhodanthe chlorocephala* ssp. *chlorocephala*, *R. manglesii*, and *R. stricta*, while an embryo dormancy which could be overcome by GA₃ limited germination of *Brachyscome iberidifolia*, *Chrysocephalum apiculatum*, *L. fitzgibbonii*, *L. molle*, *Myriocephalus stuartii*, *R. polygalifolia*, and *R. moschata* (Table 1).

Light stimulated germination of *Brachyscome iberidifolia*, *C. apiculatum*, *Hyalosperma glutinosum* ssp. *venustum*, *L. fitzgibbonii*, *L. stipitatum*, *R. humboldtiana*, *R. stricta*, and *Waitzia acuminata* (Table 2).

On the other hand, germination of *R. chlorocephala* ssp. *chlorocephala* was inhibited by light in control treatments (Table 2.).

Recommendations for seed propagation of seventeen species are given in Table 3.

CONCLUSIONS

This study has documented the germination characteristics of seventeen Asteraceae native to Australia. The pericarp and testa were influential in suppressing germination in some species, while an embryo dormancy which could be broken by GA₃ and/or light, occurred in others. It is suggested that a pre-germination treatment of GA₃ would be beneficial when germinating native daisy achenes about which little is known.

Acknowledgments. The financial support of The Horticultural Research and Development Corporation and the Australian Flora Foundation is acknowledged along with the assistance of Dot Priddy. The research was conducted at the Redlands Research Station, Ormiston.

LITERATURE CITED

- Atwater, B.R.** 1980. Germination dormancy and morphology of the seeds of herbaceous ornamental plants. *Seed Sci. and Technol.* 8:523-573.
- Bunker, K.V.** 1994. Overcoming poor germination in Australian daisies (Asteraceae) by combinations of gibberellin, scarification, light and dark. *Scientia Hort.* (in press).
- Schaumann, M.S., J. Barker, and J. Greig.** 1987. Australian daisies for gardens and floral art. Lothian, Melbourne, Australia.
- Taylorson, R.B., and S.B. Hendricks.** 1977. Dormancy in seeds. *Annu. Rev. Plant Physiol.* 28:331-354.

Table 1. Effect of scarification and GA₃ on mean percentage germination at thirty days from imbibition, of seventeen species of Australian daisies (Asteraceae) in light conditions.

Species ¹	Seed age ² (wks)	T ₅₀ ³ (days)	Germination (%)			Significance of treatment effect ⁴
			Intact (control)	Scarified	GA ₃	
<i>Brachyscome iberidifolia</i> Benth.	32	3.0	82.4a	78.7a	99.5b	**
<i>B. latisquaemea</i> F.Muell.	32	13.4	51.1ab	43.0a	64.4b	*
<i>Chrysocephalum apiculatum</i> (Labill.) Steetz. (syn. <i>Helichrysum apiculatum</i>)	28	9.8	65.9b	N ⁵	26.5a	*
<i>Hyalosperma glutinosum</i> ssp. <i>venustum</i> (Moore) Wilson (syn. <i>Helipterum venustum</i>)	28	3.4	31.7a	31.4a	37.0a	NS
<i>Leucochrysum fitzgibbonii</i> (F. Muell.) Wilson (syn. <i>Helipterum fitzgibbonii</i>)	36	5.5	18.4a	18.5a	69.6b	**
<i>L. molle</i> (Cunn. ex DC) Wilson (syn. <i>Helipterum molle</i>)	36	-	0	1.7a	17.4b+	*
<i>L. stipitatum</i> (F. Muell.) Wilson (syn. <i>Helipterum stipitatum</i>)	28	9.6	24.2a	56.3b	65.7b	**
<i>Myriocephalus stuartii</i> (F. Muell. & Sond.) Benth.	28	-	0	0	6.2	-
<i>Rhodanthe chlorocephala</i> ssp. <i>chlorocephala</i> (Turcz.) Wilson (syn. <i>Helipterum chlorocephalum</i>)	24	9.4	4.0a	17.9b	8.3 ^a	*
<i>R. chlorocephala</i> ssp. <i>rosea</i> (Hook.) Wilson (syn. <i>Helipterum roseum</i>)	24	2.0	72.4a	68.2a	78.1a	NS
<i>R. humboldtiana</i> Wilson (syn. <i>Helipterum humboldtianum</i>)	24	1.5	99.9a	99.9a	99.9a	NS
<i>R. manglesii</i> Lindley (syn. <i>Helipterum manglesii</i>)	24	2.6	38.5a	63.9b	53.1a	*
<i>R. moschata</i> (Cunn. ex DC.) Wilson (syn. <i>Helipterum moschatum</i>)	36	16.5	16.0a	22.4a	75.6b	**
<i>R. polygalifolia</i> (Cunn. ex DC.) Wilson (syn. <i>Helipterum polygalifolium</i>)	28	-	0	0	17.3	-
<i>R. stricta</i> (Lindley) Wilson (syn. <i>Helipterum stricta</i>)	28	13.4	0.5a	15.9b	31.5b	**
<i>Schoenia filifolia</i> ssp. <i>subulifolia</i> (Turcz.) Wilson (syn. <i>Helichrysum subulifolium</i>)	32	4.0	94.9a	91.3a	93.9a	NS
<i>Waitzia acuminata</i> Steetz	36	4.0	86.3a	66.2a	69.0a	NS

¹ *Chrysocephalum podolepidium* (syn. *Helichrysum podolepidium*), *Erymophyllum*

ramosum ssp. *involucratum* (syn. *Helipterum involucratum*), *Lawrencella davenportii* (syn. *Helichrysum davenportii*), *Lawrencella rosea* (syn. *Helichrysum lindeyi*), *Minuria denticulata*, *Podolepis auriculata*, *Podolepis gracilis*, *Podolepis jaceoides*, *Waitzia aurea* and *Waitzia citrina* failed to germinate during the course of the experiment.

² Seed age (weeks from collection) at beginning of germination trial.

³ T₅₀ (time to 50% germination) of intact seeds treated with water.

⁴ *, **, NS; Significant at P < 0.01, P < 0.05 and non-significant, respectively. Treatment means within rows followed by different letters, are significantly different at P < 0.05.

⁵ N = not scarified.

Table 2. Effect of GA₃, scarification and light on mean percentage germination at fifteen days from imbibition of seventeen species of Australian native daisies (Asteraceae).

Species	Germination (%)						Significant effects ¹		
	Intact		Scarified		GA ₃		light (l)	treatment (t)	t x l
	light	dark	light	dark	light	dark			
<i>Brachyscome</i>									
<i>iberidifolia</i>	82.4b	80.7b	78.7b	58.8a	99.5c	90.3b	*	**	NS
<i>B. latisquaemea</i>	28.0abc	26.2ab	13.6a	31.9bc	47.3cd	52.0d	NS	**	NS
<i>Chrysocephalum</i>									
<i>apiculatum</i>	56.9c	0.05a	N ²	N ²	24.7b	17.1b	**	NS	**
<i>Hyalosperma</i>									
<i>glutinosum</i>									
ssp. <i>venustum</i>	31.7b	0.05a	30.0b	0.6a	31.0b	34.4b	**	**	**
<i>Leucochrysum</i>									
<i>fitgibbonnu</i>	18.4b	1.7a	16.2b	1.7a	62.3d	40.9c	**	**	NS
<i>L. molle</i>	0	0	1.7a	6.3ab+	15.3b+	6.1a+	NS	*	*
<i>L. stipitatum</i>	15.4b	1.4a	25.7bc	0.05a	48.5d	41.4cd	**	**	*
<i>Myriocephalus</i>									
<i>stuartii</i>	0	0	0	0	4.4a+	9.4a+	-	NS	-
<i>Rhodanthe</i>									
<i>chlorocephalasp.</i>									
<i>chlorocephala</i>	1.4a	27.3c	11.7bc	6.5ab	8.3ab	3.7a	NS	NS	**
<i>R. chlorocephala</i>									
ssp. <i>rosea</i>	72.4a	62.6a	66.9a	62.8a	76.9a	97.7b	NS	**	*
<i>R. humboldtiana</i>	99.9b	99.9b	99.9b	99.9b	99.9b	97.1a	**	**	**
<i>R. manglesii</i>	37.1a	37.5a	68.4cd	77.7d	48.6ab	58.0bc	NS	**	NS
<i>R. moschata</i>	5.3a	18.5ab	11.2a	37.9bc	66.0d	59.1cd	NS	**	NS
<i>R. polygalifolia</i>	0	0	0	0	4.3a+	12.0a+	-	NS	-
<i>R. stricta</i>	0.5a+	0.05a	14.6b+	4.2ab+	12.4b+	1.4a	**	**	NS
<i>Schoenia filifolia</i>									
ssp. <i>subulifolia</i>	94.9a	89.8a	86.2a	86.7a	93.9a	91.6a	NS	NS	NS
<i>Waitzia</i>									
<i>acuminata</i>	34.5b	7.4a	24.8b	3.1a	69.0c	54.8c	**	**	NS

¹ *, **, NS; Significant at $P < 0.01$, $P < 0.05$ and nonsignificant respectively; +, significantly different from zero. Treatment means in the same row followed by different letters are significantly different at $P < 0.05$.

² N = not scarified.

Table 3. Recommendations for seed propagation of seventeen species of Australian daisies.

Species	Pre-sowing treatment	Light response	Sowing
<i>Brachyscome iberidifolia</i>	GA ₃	positive	surface
<i>B. latisqaemea</i>	no	neutral	surface/shallow
<i>Chrysocephalum apiculatum</i>	no	positive	surface
<i>Hyalosperma glutinosum</i> ssp. <i>venustum</i>	no	positive	surface
<i>Leucochrysum fitzgibbonii</i>	GA ₃	positive	surface
<i>L. molle</i>	GA ₃	neutral	surface/shallow
<i>L. stipitatum</i>	scarify or GA ₃	positive	surface
<i>Myriocephalus stuartii</i>	GA ₃	neutral	surface/shallow
<i>Rhodanthe chlorocephala</i> ssp. <i>chlorocephala</i>	scarify	negative	covered
<i>R. chlorocephala</i> ssp. <i>rosea</i>	no	neutral	surface/shallow
<i>R. humboldtiana</i>	no	positive	surface
<i>R. manglesii</i>	scarify	neutral	surface/shallow
<i>R. moschata</i>	GA ₃	neutral	surface/shallow
<i>R. polygalifolia</i>	GA ₃	neutral	surface/shallow
<i>R. stricta</i>	scarify or GA ₃	positive	surface
<i>Schoenia filifolia</i> ssp. <i>subulifolia</i>	no	neutral	surface/shallow
<i>Waitzia acuminata</i>	no	positive	surface